

DOI: <https://doi.org/10.17816/aog633897>

Bacterial vaginosis biofilms: a target for therapeutic innovation

Kseniya A. Rossolovskaya, Natalia S. Trifonova, Irina V. Gadaeva, Leonid G. Spivak

I.M. Sechenov First Moscow State Medical University, Moscow, Russia

ABSTRACT

Bacterial vaginosis (BV) is one of the most common vaginal microbiome abnormalities worldwide and a risk factor for various obstetric and gynecological complications.

Despite years of exploration, existing and quickly emerging clinical, laboratory and instrumental diagnostic methods, and progressive development of science in general, the etiology and pathogenesis of BV remain poorly understood. This is evidenced by the high incidence of chronic and/or recurrent course. There are standard therapeutic approaches aimed to eradicating the causative agent, but the level of efficacy remains questionable due to recurrent episodes. Therefore, further studies of this problem are warranted. Actually, it is evident that *G. vaginalis* forms polymicrobial biofilms on urogenital tract mucosa.

Biofilms represent associations of microorganisms that are adhered to the surface of the epithelium and connected together in the polymer matrix. Biofilms change the properties of the microorganisms involved into their structural frame and provide beneficial conditions for their interactions. This results in the increase of the existing pathogenic properties of bacteria associated with BV, as well as in the appearance of new features. Thus, the microorganisms become less susceptible to previously effective antibiotics and to aggressive media. Finally, this contributes to the recurrent course of the disease.

In most cases, treatment of BV is based on the immediate effect on the microorganisms, but in patients with confirmed biofilm-associated BV this strategy is not effective and is associated with BV recurrences. Thus, currently relevant issues include exploration of the causes of recurrent BV, development of anti-biofilm agents able to disrupt their matrix and release bacteria from their carcass, and introduction of these agents into clinical practice. This will increase the effectiveness of treatment.

Keywords: bacterial vaginosis; etiology; biofilms; treatment.

To cite this article:

Rossolovskaya KA, Trifonova NS, Gadaeva IV, Spivak LG. Bacterial vaginosis biofilms: a target for therapeutic innovation. *V.F. Snegirev Archives of Obstetrics and Gynecology*. 2024;11(4):406–415. DOI: <https://doi.org/10.17816/aog633897>

Received: 27.06.2024

Accepted: 29.07.2024

Published online: 12.12.2024

DOI: <https://doi.org/10.17816/aog633897>

Биоплёнки бактериального вагиноза — мишень для терапевтического новаторства

К.А. Россоловская, Н.С. Трифонова, И.В. Гадаева, Л.Г. Спивак

Первый Московский государственный медицинский университет им. И.М. Сеченова, Москва, Россия

АННОТАЦИЯ

Бактериальный вагиноз — один из наиболее распространённых вариантов нарушения вагинального микробиома во всём мире. Он является фактором риска развития различных осложнений в акушерской и гинекологической практике.

Несмотря на многолетнее изучение этого синдрома, существующие и динамично совершенствующиеся клинико-лабораторные и инструментальные методы диагностики, прогрессивное развитие науки в целом, этиология и патогенез БВ до сих пор недостаточно изучены. Об этом свидетельствует высокая частота хронического и/или рецидивирующего течения. Существуют стандартные терапевтические подходы, направленные на эрадикацию этиологического агента, однако уровень эффективности остаётся сомнительным из-за повторяющихся эпизодов, что требует дальнейшего изучения данной проблемы. В настоящее время достоверно установлено, что *G. vaginalis* формирует на слизистой урогенитального тракта полимикробные биоплёнки.

Биоплёнка представляет собой кооперацию микроорганизмов, которые прикреплены к поверхности эпителия и связаны между собой полимерным матриксом. Биоплёнка оказывает влияние на свойства микроорганизмов, входящих в её структуру, обеспечивает условия для их взаимодействия. Это приводит не только к усилению уже имеющихся патогенных свойств бактерий, ассоциированных с бактериальным вагинозом, но и к формированию новых характеристик, благодаря которым микроорганизмы становятся в меньшей степени восприимчивы к ранее эффективным антибактериальным препаратам и агрессивным средам, что способствует рецидивированию.

В большинстве случаев лечение бактериального вагиноза осуществляется по прямому механизму воздействия на микроорганизмы, однако у пациенток с верифицированным биоплёночным бактериальным вагинозом такая стратегия нерезультативна, что приводит к рецидивам. Таким образом, на сегодняшний день остаются актуальными диагностика причин рецидивирующего бактериального вагиноза, разработка и внедрение в клиническую практику антибиоплёночных агентов, нацеленных на разрушение матрикса биоплёнки и освобождение из его структуры бактерий, что позволит повысить эффективность лечения.

Ключевые слова: бактериальный вагиноз; этиология; биоплёнки; лечение.

Как цитировать:

Россоловская К.А., Трифонова Н.С., Гадаева И.В., Спивак Л.Г. Биоплёнки бактериального вагиноза — мишень для терапевтического новаторства // Архив акушерства и гинекологии им. В.Ф. Снегирёва. 2024. Т. 11, № 4. С. 406–415. DOI: <https://doi.org/10.17816/aog633897>

DOI: <https://doi.org/10.17816/aog633897>

细菌性阴道病生物膜：治疗创新的靶点

Kseniya A. Rossolovskaya, Natalia S. Trifonova, Irina V. Gadaeva, Leonid G. Spivak

I.M. Sechenov First Moscow State Medical University, Moscow, Russia

摘要

细菌性阴道病（Bacterial Vaginosis, BV）是全球范围内最常见的阴道微生物群失调类型之一，同时是产科和妇科实践中各种并发症的重要风险因素。

尽管这一综合征已被研究多年，并且现有的临床实验室和仪器诊断方法不断完善，但BV的病因和发病机制仍未完全阐明。这可以从其高慢性化和高复发率的特点中看出。目前的标准治疗方法主要旨在根除病原体，但由于复发率高，治疗效果仍然有限，这需要进一步深入研究。研究表明，*G. vaginalis* 在尿生殖道黏膜上形成多菌种生物膜。

生物膜是微生物的集合体，其附着在上皮表面，并通过聚合物基质相互连接。生物膜改变了其内部微生物的特性，为其提供相互作用的条件。这不仅增强了与BV相关的细菌既有的致病特性，还赋予了它们新的特性，包括降低对以往有效抗菌药物和不利环境的敏感性，从而导致复发。

在大多数情况下，BV的治疗直接针对微生物，但对于被诊断为生物膜型BV的患者，这种治疗策略效果有限，往往导致复发。因此，目前的研究重点是明确复发性BV的原因，并开发针对生物膜基质破坏的抗生物膜药物。这些药物能够释放基质内的细菌，从而显著提高治疗效果。

关键词：细菌性阴道病；病因学；生物膜；治疗。

引用本文：

Rossolovskaya KA, Trifonova NS, Gadaeva IV, Spivak LG. 细菌性阴道病生物膜：治疗创新的靶点. *V.F. Snegirev Archives of Obstetrics and Gynecology*. 2024;11(4):406–415. DOI: <https://doi.org/10.17816/aog633897>

收到: 27.06.2024

接受: 29.07.2024

发布日期: 12.12.2024

BACKGROUND

Bacterial vaginosis (BV) is one of the most common gynecological conditions that does not directly affect a woman's health, but significantly impairs her quality of life [1]. Complications in patients with BV can be both gynecological (pelvic inflammatory disease, infertility of various origins, etc.) and obstetric (in women with BV, rates of miscarriage and preterm delivery are 3–5 and 2–7 fold higher, respectively, depending on gestational age) [2–4]. These complications can seriously affect women's reproductive health and therefore require attention of the medical community.

The global literature estimates the prevalence of BV in the female population to be approximately 29%, or more than 21 million diagnosed patients [5]. The prevalence of BV varies by geographic region and ethnicity. For example, a systematic review by Kenyon et al. [6] showed that the highest BV prevalence was reported in some parts of Africa (Botswana, Central African Republic, Gambia, Ghana, etc.), while most countries in Asia and Europe had low levels of the syndrome. A systematic review and meta-analysis by Peebles et al. found both regional and racial differences in the global prevalence of BV; rates of BV in African American and Hispanic women (33% and 31%, respectively) are significantly higher than in European (23%) and Asian (11%) women [7]. BV is a serious issue for women of childbearing age (23%–29%), which may affect the overall demographics, as the health of this population is directly related to the birth rate and the health of future generations [8]. In addition, the incidence of chronic and/or recurrent BV is increasing and the cost of diagnosis and treatment is a significant burden on the global economy [4]. For example, a meta-analysis by Peebles et al. [9] showed that annual cost burden of BV treatment is high, at an estimated US \$4.8 billion.

BV can present in various clinical forms; one in four patients is asymptomatic, while symptomatic BV is often accompanied by a homogeneous gray or grayish-white foul-smelling (fishy odor) vaginal discharge, itching, burning, and other vaginal discomfort [5, 10]. Diagnosis of BV includes conventional modalities: vaginal pH, which is usually elevated in BV; the amine test (addition of 10% potassium hydroxide to vaginal discharge will produce a characteristic fishy odor); microscopy to detect key cells relevant to BV; microbiology to determine the composition of the microflora. New modalities are also being used, such as polymerase chain reaction for qualitative and quantitative assessment of vaginal bacterial composition, biochemical or molecular biotyping, and high-throughput sequencing, which provide a more comprehensive picture of microbiome composition [11].

HISTORY OF BACTERIAL VAGINOSIS RESEARCH

In practice, BV is usually characterized as a polymicrobial non-inflammatory condition caused by the decreased

proportion of *Lactobacilli* or their complete absence in the vaginal microbiome and increased counts of obligate and facultative anaerobic opportunistic pathogens [12]. The vaginal microbiome is a complex, dynamic ecosystem that fluctuates throughout a menstrual period and an entire woman's life. Some authors report the bacterial count in young women of childbearing age to be 10^{10} – 10^{11} CFU/mL [13]. The vaginal microbiome of a healthy woman is maintained in homeostasis. However, exogenous and endogenous factors may affect this balance. Endogenous factors include alterations in hormone levels, a woman's age, weight, pregnancy, immune status, etc. Exogenous factors include the use of antibacterial agents, genital/extragenital infections, and inflammation [14]. *Lactobacilli* are the main bacteria that maintain a stable and healthy vaginal microbiocenosis. Albert Döderlein [15] was the first who discovered rod-shaped bacilli in the vagina and cultured them in a nutrient broth. He also found that these microorganisms are capable of producing lactic acid in the vagina, making it acidic, and proposed the concept of "vaginal acidity."

The history of vaginal microbiome research dates back to the mid-19th century, when clinicians noticed increasing rates of obstetrical and gynecological conditions caused by non-compliance with aseptic and antiseptic practice. The clinical use of light microscopy allowed in-depth research into the vaginal microflora composition and the search for causes of infectious complications in women. The identification of an etiologic agent and the development of a taxonomic classification of microbes associated with BV have been a challenge for many years due to the high selectivity of *Gardnerella vaginalis* to growth media, its unique physical and chemical structure and molecular architecture of the cell wall, genome diversity, and unclear pathogenic factors. For example, in 1953, Leopold published a study evaluating cervical discharge and urine in women with cervicitis and men with prostatitis, respectively. He detected a rod-shaped bacterium, which he described as small, gram-negative, non-motile, unencapsulated. Leopold could not identify the species of this bacterium, but based on its morphological characteristics, he proposed that it belonged to *Haemophilus* [16]. The microorganism previously described by Leopold was later discovered in 1955 by Gardner and Dukes [17] in women with non-specific vaginitis. Due to its tropism for blood-containing media, this rod-shaped bacterium was also classified as a member of the genus *Haemophilus* and named *Haemophilus vaginalis* due to its location. Since then, the condition associated with this pathogen has been classified as *Haemophilus vaginalis vaginitis*.

The species identification for the microorganism described by Leopold, Gardner, and Dukes has been the focus of research for many scientists since the 1960s, as subsequent studies provided conflicting results. In 1963, Zinnemann and Turnerg proposed to classify *Haemophilus vaginalis* as a member of the genus *Corynebacterium* and named it

Corynebacterium vaginale [18]. The taxonomic classification did not include the microorganism in *Corynebacteria*. *Gardnerella vaginalis*, formerly known as *Corynebacterium vaginale* and *Haemophilus vaginalis*, was named after Hermann L. Gardner, who discovered and classified it as the only member of the genus *Gardnerella* [19, 20]. Later, some features of *Gardnerella vaginalis* were identified: it was a gram-variable, pleomorphic, non-motile, non-sporeforming, acapsular, flagellar bacterium ranging from 0.4–1.5 μm to 2–3 μm [21]. In addition, the size and morphology of the cells vary significantly depending on growth conditions and physiological status [22]. Due to the unique structure, *Gardnerella vaginalis* has attracted not only clinicians but also basic scientists, driving ongoing research in the field. Electron microscopy revealed fimbriae on the *Gardnerella vaginalis* surface. The fimbriae mediate adhesion to vaginal epithelium *in vivo* and to desquamated epithelium, which are the key cells [23]. Microscopy and cultivation of *Gardnerella vaginalis* often found that its cells were often linked together, which was explained by the production of exopolysaccharide with bacteria binding its filaments together [24]. It is now recognized that this is a biofilm consisting of microorganisms bound by a polymer matrix [25].

BIOFILMS: PATHOGENESIS OF BACTERIAL VAGINOSIS

Currently, biofilm-associated BV is detected in 90% of patients [8]. Of note that this term, as well as the term “biofilm-associated gardnerellosis” does not exist officially, but it is proposed by some authors to clarify the etiopathogenesis of the disease. Biofilms are a structured community of one or more species embedded in a polymer matrix consisting of proteins, carbohydrates, and nucleic acids [26, 27]. The biofilm concentration of some microbes can reach 10^{11} CFU/mL [28]. The American researcher John William Costerton described this phenomenon and introduced the term “biofilm” [29]. The formation of biofilms is a complex, stepwise, and dynamic process, as well as an effective survival strategy of microorganisms under unfavorable conditions [30]. Biofilm microorganisms are capable of regulating the production of virulence factors through an identified type of communication called quorum sensing. The biofilm cycle includes three stages: adhesion to the surface of the vaginal epithelium, secretion of the polymer matrix, and aggregation of microbes on the surface of the mature biofilm [27]. Biofilms have a higher tolerance to the aggressive factors compared to free-living microorganisms in the vaginal tract. This is possible due to the unique multi-component biofilm structure and the layered organization of microorganisms within the biofilm [31]. This structure contributes to the impaired diffusion of antimicrobial agents through the biofilm, reduced metabolic activity of the cells, and the emergence of antibiotic-resistant bacteria [32, 33].

For a long time, the role of *Gardnerella* in the biofilm formation was unclear: does *Gardnerella vaginalis* trigger this process or does it support the matrix of the biofilm formed by the association of bacteria? However, current literature suggests that *G. vaginalis* is a major colonizer that can provide a matrix for attachment of other BV-associated microorganisms, thereby enabling the formation of polymicrobial biofilms [27]. Some researchers, such as Bonnardel et al. [34], linked the mechanism of biofilm formation to bacterial lectins of *Gardnerella*, which are tropic to glycosylated components of the mucosa, thus forming a vaginal cell–*Gardnerella* complex. A study by Martin reported a role of a collagen-binding protein produced by *Gardnerella*, which may also contribute to the biofilm formation and possibly immune evasion through interaction with complement proteins [35]. In addition, regarding the microbiological features of *Gardnerella*, its significant adhesion to vaginal cells and ability to form biofilms confirm the trigger role of *Gardnerella* in the colonization of the vaginal epithelium, acting as a frame for the adherence of other species. Modern molecular diagnostic modalities show that *G. vaginalis* constitutes 60–90% of the biofilm. Other common biofilm members include *Sneathia sanguinegens*, *Porphyromonas assaccharolytica*, *Megasphaera* spp. and the difficult-to-culture *Atopobium vaginae*, which may comprise up to 40% of the biofilm [5]. Their role in vaginal biofilm formation and pathogenicity of *G. vaginalis* has not been adequately evaluated [36]. Some studies reported that some, but not all, types of vaginal microflora can participate in biofilm formation and enhance *G. vaginalis* virulence [37]. Other studies found that microorganisms other than *G. vaginalis* may increase total biofilm biovolume [38]. Therefore, biofilm formation and development depend on both the triggering effect of *Gardnerella* and other vaginal microorganisms that form a kind of social network and thereby regulate the pathogenetic process of biofilm-associated BV. As a result, BV involves a complex interaction between opportunistic pathogens, endogenous vaginal microbiota, and the vaginal epithelium [39].

The main biofilm component is an extracellular polysaccharide matrix (exopolysaccharide), which constitutes 85% of its volume [25]. The matrix is produced by, and is composed of, the bacteria associated with BV, and it organizes the biofilm structurally and protects it from physical, chemical, and immune factors [40–42]. Nowadays, biofilms are thought to contribute to recurrent BV, although the term “recurrent BV” does not currently have a generally accepted definition [41]. There is no classification of BV in the relevant clinical guidelines published in 2022. The literature describes recurrent BV in a variety of ways. For example, Marshall et al. [43] considered this diagnosis in patients who “recur one or more times after completion of an episodic regimen,” Letyaeva [1] diagnosed recurrent BV in patients with more than four episodes per year, and Pestrikova et al. used the criterion of 3–4 episodes per year [28].

CONCLUSION

There is an increasing research focus on biofilm as a pathogenetic factor of BV. Most treatment options focus on the etiologic agent of a disease, i.e. the direct mechanism of action. However, the ability of microorganisms to adapt to the effects of aggressive agents makes many treatment regimens ineffective, as evidenced by increased recurrence rates. The data on the structure of the biofilm confirmed the importance of the development and clinical use of agents that disrupt the polysaccharide matrix of the biofilm and release the bacteria present in this matrix (dispersants), and this is the focus of our research. This will both improve treatment outcomes and enable the practical use of previously effective and established treatment options.

ADDITIONAL INFO

Authors' contributions. K.A. Rossolovskaya performed literature review, collected and analyzed literature sources, prepared, wrote and edited the manuscript; N.S. Trifonova reviewed and edited the manuscript; I.V. Gadaeva reviewed and edited the manuscript; L.G. Spivak reviewed and edited the manuscript. All authors confirm that their authorship meets the international ICMJE criteria (all authors have made a significant contribution to the development of the concept, research and preparation of the article, read and approved the final version before publication).

Funding source. This study was not supported by any external sources of funding.

Competing interests. The authors declares that there are no obvious and potential conflicts of interest associated with the publication of this article.

REFERENCES

1. Letyaeva OI. Bacterial vaginosis: current opportunities and prospects for long-term control. *Russian Bulletin of Obstetrician-Gynecologist*. 2019;19(2):100–104. EDN: FHTCEF doi: 10.17116/rosakush201919021100
2. Tabatabaei N, Eren AM, Barreiro LB, et al. Vaginal microbiome in early pregnancy and subsequent risk of spontaneous preterm birth: a case-control study. *BJOG*. 2019;126(3):349–358. doi: 10.1111/1471-0528.15299
3. Johnston W, Ware A, Kuiters WF, et al. *In vitro* bacterial vaginosis biofilm community manipulation using endolysin therapy. *Biofilm*. 2022;5:100101. doi: 10.1016/j.biofilm.2022.100101
4. Bretelle F, Loubière S, Desbrière R, et al. Effectiveness and costs of molecular screening and treatment for bacterial vaginosis to prevent preterm birth: The AuTop randomized clinical trial. *JAMA Pediatr*. 2023;177(9):894–902. doi: 10.1001/jamapediatrics.2023.2250
5. Novikova SV, Tsvitsivadze EB, Fedotova AV. Bacterial vaginosis as a typical biofilm infection. *Russian Bulletin of Obstetrician-Gynecologist*. 2018;18(4):97–100. EDN: XWAUCT doi: 10.17116/rosakush201818497
6. Kenyon C, Colebunders R, Crucitti T. The global epidemiology of bacterial vaginosis: a systematic review. *Am J Obstet Gynecol*. 2013;209(6):505–523. doi: 10.1016/j.ajog.2013.05.006
7. Peebles K, Velloza J, Balkus JE, et al. High global burden and costs of bacterial vaginosis: a systematic review and meta-analysis. *Sex Transm Dis*. 2019;46(5):304–311. doi: 10.1097/OLQ.0000000000000972
8. Dobrokhotova YuE, Kazantseva VD, Bondarenko KR. Bacterial vaginosis: modern anti-relapse treatment tactics. *RMJ*. 2022;(8):61–65. EDN: GVQLAZ
9. Javed A, Parvaiz F, Manzoor S. Bacterial vaginosis: An insight into the prevalence, alternative treatments regimen and its associated resistance patterns. *Microb Pathog*. 2019;127:21–30. doi: 10.1016/j.micpath.2018.11.046
10. Reiter S, Kellogg Spadt S. Bacterial vaginosis: a primer for clinicians. *Postgrad Med*. 2019;131(1):8–18. doi: 10.1080/00325481.2019.1546534
11. Muzny CA, Cerca N, Elnaggar JH, et al. State of the Art for Diagnosis of Bacterial Vaginosis. *J Clin Microbiol*. 2023;61(8):e0083722. doi: 10.1128/jcm.00837-22
12. Bacterial vaginosis: Clinical recommendations under. Moscow, 2022 [cited 2024 Jun 02]. Available from: https://cr.minzdrav.gov.ru/recomend/206_2 (In Russ.)
13. Chen C, Song X, Wei W, et al. The microbiota continuum along the female reproductive tract and its relation to uterine-related diseases. *Nat Commun*. 2017;8(1):875. doi: 10.1038/s41467-017-00901-0
14. Pekmezovic M, Mogavero S, Naglik JR, Hube B. Host-pathogen interactions during female genital tract infections. *Trends Microbiol*. 2019;27(12):982–996. doi: 10.1016/j.tim.2019.07.006
15. Döderlein A. Das scheidensekret und seine bedeutung für das puerperalfieber. Leipzig: Verlag von Eduard Besold; 1892.
16. Leopold S. Heretofore undescribed organism isolated from the genitourinary system. *US Armed Forces Med*. 1953;4(2):263–266.
17. Gardner HL, Dukes CD. *Haemophilus vaginalis* vaginitis: a newly defined specific infection previously classified non-specific vaginitis. *Am J Obstet Gynecol*. 1955;69(5):962–976.
18. Zinnemann K, Turnerg C. The taxonomic position of 'Haemophilus vaginalis' (*Corynebacterium vaginale*). *J Pathol Bacteriol*. 1963;85(1):213–219. doi: 10.1002/PATH.1700850120
19. Greenwood JR, Pickett MJ. Transfer of *Haemophilus vaginalis* Gardner and Dukes to a new genus, *Gardnerella*: *G. vaginalis* (Gardner and Dukes). *Int J Syst Bacteriol*. 1980;30(1):170–178. doi: 10.1099/00207713-30-1-170
20. Piot P, van Dyck E, Goodfellow M, Falkow S. A taxonomic study of *Gardnerella vaginalis* (*Haemophilus vaginalis*) Gardner and Dukes 1955. *J Gen Microbiol*. 1980;119(2):373–396. doi: 10.1099/00221287-119-2-373
21. Piot P. *Gardnerella*, *streptobacillus*, *spirillum*, and *calymmatobacterium*. In: Balows A, Hausler WJ Jr, Herrmann KL, Isenberg HD, Shadomy HJ, editors. *Manual of Clinical Microbiology*. 5th ed. Washington, D.C: American Society for Microbiology; 1991:483–487.
22. Sadhu K, Domingue PA, Chow AW, et al. *Gardnerella vaginalis* has a Gram-positive cell-wall ultrastructure and lacks classical cell-wall lipopolysaccharide. *J. Med. Microbiol*. 1989;29(3):229–235. doi: 10.1099/00222615-29-3-229
23. Scott TG, Curran B, Smyth CJ. Electron microscopy of adhesive interactions between *Gardnerella vaginalis* and vaginal epithelial cells, McCoy cells and human red blood cells. *J Gen Microbiol*. 1989;135(3):475–480. doi: 10.1099/00221287-135-3-475
24. Taylor-Robinson D. The bacteriology of *Gardnerella vaginalis*. *Scand J Urol. Nephrol Suppl*. 1984;86:41–55.
25. Ilyina TS, Romanova YuM. The role of bacterial biofilms in chronic infectious processes and the search for methods to combat them. *Molecular Genetics, Microbiology and Virology*. 2021;39(2):14–24. EDN: RHLJAM doi: 10.17116/molgen20213902114
26. Khryanin AA. Microbial biofilms: modern concepts. *Antibiotics and Chemotherapy*. 2020;65(5–6):70–77. EDN: NQITOE doi: 10.37489/0235-2990-2020-65-5-6-70-77
27. Jung HS, Ehlers MM, Lombaard H, et al. Etiology of bacterial vaginosis and polymicrobial biofilm formation. *Crit Rev Microbiol*. 2017;43(6):651–667. doi: 10.1080/1040841X.2017.1291579
28. Pestrikova TYu, Yurasova EA, Kotelnikova AV, et al. Modern approach to treatment of a recurrent bacterial vaginosis at women of the reproductive period. *Gynecology*. 2018;20(2):55–58. EDN: XTGRVB doi: 10.26442/2079-5696_2018.2.55-58
29. Nickel JC, Ruseska I, Wright JB, Costerton JW. Tobramycin resistance of cells of *Pseudomonas aeruginosa* growing as a biofilm on urinary catheter material. *Antimicrob Agents Chemother*. 1985;27(4):619–624. doi: 10.1128/AAC.27.4.619
30. Berezovskaya ES, Makarov IO, Gornberg MA, et al. Biofilm formation at the bacterial vaginosis. *Obstetrics, Gynecology and Reproduction*. 2013;7(2):34–36. EDN: RRPOER
31. Simões M, Simões LC, Vieira MJ. Species association increases biofilm resistance to chemical and mechanical treatments. *Water Res*. 2009;43(1):229–237. doi: 10.1016/j.watres.2008.10.010

32. Khan J, Tarar SM, Gul I, et al. Challenges of antibiotic resistance biofilms and potential combating strategies: a review. *3 Biotech*. 2021;11(4):169. doi: 10.1007/s13205-021-02707-w
33. Michaelis C, Grohmann E. Horizontal gene transfer of antibiotic resistance genes in biofilms. *Antibiotics (Basel)*. 2023;12(2):328. doi: 10.3390/antibiotics12020328
34. Bonnardel F, Haslam SM, Dell A, et al. Proteome-wide prediction of bacterial carbohydrate-binding proteins as a tool for understanding commensal and pathogen colonisation of the vaginal microbiome. *NPJ Biofilms Microbiomes*. 2021;7(1):49. doi: 10.1038/s41522-021-00220-9
35. Marín E, Haesaert A, Padilla L, et al. Unraveling *Gardnerella vaginalis* surface proteins using cell shaving proteomics. *Front Microbiol*. 2018;9:975. doi: 10.3389/fmicb.2018.00975
36. Hardy L, Jespers V, Abdellati S, et al. A fruitful alliance: the synergy between *Atopobium vaginae* and *Gardnerella vaginalis* in bacterial vaginosis-associated biofilm. *Sex Transm Infect*. 2016;92(7):487–491. doi: 10.1136/sextrans-2015-052475
37. Castro J, Machado D, Cerca N. Unveiling the role of *Gardnerella vaginalis* in polymicrobial Bacterial Vaginosis biofilms: the impact of other vaginal pathogens living as neighbors. *ISME J*. 2019;13(5):1306–1317. doi: 10.1038/s41396-018-0337-0
38. Castro J, Cerca N. BV and non-BV associated *Gardnerella vaginalis* establish similar synergistic interactions with other BV-associated microorganisms in dual-species biofilms. *Anaerobe*. 2015;36:56–59. doi: 10.1016/j.anaerobe.2015.10.008
39. Schwebke JR, Muzny CA, Josey WE. Role of *Gardnerella vaginalis* in the pathogenesis of bacterial vaginosis: a conceptual model. *J Infect Dis*. 2014;210(3):338–343. doi: 10.1093/infdis/jiu089
40. Shvartsman E, Hill JE, Sandstrom P, MacDonald KS. *Gardnerella* revisited: species heterogeneity, virulence factors, mucosal immune responses, and contributions to bacterial vaginosis. *Infect Immun*. 2023;91(5):e0039022. doi: 10.1128/iai.00390-22
41. Coudray MS, Madhivanan P. Bacterial vaginosis — A brief synopsis of the literature. *Eur J Obstet Gynecol Reprod Biol*. 2020;245:143–148. doi: 10.1016/j.ejogrb.2019.12.035
42. Abaturov AE. Polysaccharide-degrading enzymes as agents dispersing bacterial biofilms. *Zdorov'e Rebenka*. 2020;15(4):271–278. EDN: WKPGMH doi: 10.22141/2224-0551.15.4.2020.208478
43. Marshall AO. Managing recurrent bacterial vaginosis: insights for busy providers. *Sex Med Rev*. 2015;3(2):88–92. doi: 10.1002/smrj.45

СПИСОК ЛИТЕРАТУРЫ

1. Летяева О.И. Бактериальный вагиноз: современные возможности и перспективы длительного контроля // Российский вестник акушера-гинеколога. 2019. Т. 19, № 2. С. 100–104. EDN: FHTCEF doi: 10.17116/rosakush201919021100
2. Tabatabaei N., Eren A.M., Barreiro L.B., et al. Vaginal microbiome in early pregnancy and subsequent risk of spontaneous preterm birth: a case-control study // BJOG. 2019. Vol. 126, N 3. P. 349–358. doi: 10.1111/1471-0528.15299
3. Johnston W., Ware A., Kuiters W.F., et al. *In vitro* bacterial vaginosis biofilm community manipulation using endolysin therapy // Biofilm. 2022. Vol. 5. P. 100101. doi: 10.1016/j.bioflm.2022.100101
4. Bretelle F., Loubière S., Desbriere R., et al. Effectiveness and costs of molecular screening and treatment for bacterial vaginosis to prevent preterm birth: The AuTop randomized clinical trial // JAMA Pediatr. 2023. Vol. 177, N 9. P. 894–902. doi: 10.1001/jamapediatrics.2023.2250
5. Новикова С.В., Цивцивадзе Е.Б., Федотова А.В. Бактериальный вагиноз как типичная биопленочная инфекция // Российский вестник акушера-гинеколога. 2018. Т. 18, № 4. С. 97–100. EDN: XWAUCT doi: 10.17116/rosakush201818497
6. Kenyon C., Colebunders R., Crucitti T. The global epidemiology of bacterial vaginosis: a systematic review // Am J Obstet Gynecol. 2013. Vol. 209, N 6. P. 505–523. doi: 10.1016/j.ajog.2013.05.006
7. Peebles K., Velloza J., Balkus J.E., et al. High global burden and costs of bacterial vaginosis: a systematic review and meta-analysis // Sex Transm Dis. 2019. Vol. 46, N 5. P. 304–311. doi: 10.1097/OLQ.0000000000000972
8. Доброхотова Ю.Э., Казанцева В.Д., Бондаренко К.Р. Бактериальный вагиноз: современные противорецидивные стратегии // РМЖ. 2022. № 8. С. 61–65. EDN: GVQLAZ
9. Javed A., Parvaiz F., Manzoor S. Bacterial vaginosis: An insight into the prevalence, alternative treatments regimen and its associated resistance patterns // Microb Pathog. 2019. Vol. 127. P. 21–30. doi: 10.1016/j.micpath.2018.11.046
10. Reiter S., Kellogg Spadt S. Bacterial vaginosis: a primer for clinicians // Postgrad Med. 2019. Vol. 131, N 1. P. 8–18. doi: 10.1080/00325481.2019.1546534
11. Muzny C.A., Cerca N., Elnaggar J.H., et al. State of the art for diagnosis of bacterial vaginosis // J Clin Microbiol. 2023. Vol. 61, N 8. e0083722. doi: 10.1128/jcm.00837-22
12. Бактериальный вагиноз: Клинические рекомендации. Москва, 2022. Режим доступа: https://cr.minzdrav.gov.ru/recomend/206_2 Дата обращения: 02.06.2024.
13. Chen C., Song X., Wei W., et al. The microbiota continuum along the female reproductive tract and its relation to uterine-related diseases // Nat Commun. 2017. Vol. 8, N 1. P. 875. doi: 10.1038/s41467-017-00901-0
14. Pekmezovic M., Mogavero S., Naglik J.R., Hube B. Host-pathogen interactions during female genital tract infections // Trends Microbiol. 2019. Vol. 27, N 12. P. 982–996. doi: 10.1016/j.tim.2019.07.006
15. Döderlein A. Das scheidensekret und seine bedeutung für das puerperalfieber. Leipzig: Verlag von Eduard Besold, 1892.
16. Leopold S. Heretofore undescribed organism isolated from the genitourinary system // US Armed Forces Med. 1953. Vol. 4, N 2. P. 263–266.
17. Gardner H.L., Dukes C.D. *Haemophilus vaginalis* vaginitis: a newly defined specific infection previously classified non-specific vaginitis // Am J Obstet Gynecol. 1955. Vol. 69, N 5. P. 962–976.
18. Zinnemann K., Turnerg C. The taxonomic position of 'Haemophilus vaginalis' (*Corynebacterium vaginale*) // J Pathol Bacteriol. 1963. Vol. 85, N 1. P. 213–219. doi: 10.1002/PATH.1700850120
19. Greenwood J.R., Pickett M.J. Transfer of *Haemophilus vaginalis* Gardner and Dukes to a new genus, *Gardnerella*: *G. vaginalis* (Gardner and Dukes) // Int J Syst Bacteriol. 1980. Vol. 30, N 1. P. 170–178. doi: 10.1099/00207713-30-1-170

20. Piot P., van Dyck E., Goodfellow M., Falkow S. A taxonomic study of *Gardnerella vaginalis* (*Haemophilus vaginalis*) Gardner and Dukes 1955 // *J Gen Microbiol.* 1980. Vol. 119, N 2. P. 373–396. doi: 10.1099/00221287-119-2-373
21. Piot P. *Gardnerella*, *streptobacillus*, *spirillum*, and *calymmatobacterium*. In: Balows A., Hausler W.J. Jr, Herrmann K.L., Isenberg H.D., Shadomy H.J., editors. *Manual of Clinical Microbiology*. 5th ed. Washington, D.C: American Society for Microbiology; 1991:483–487.
22. Sadhu K., Domingue P.A., Chow A.W., et al. *Gardnerella vaginalis* has a Gram-positive cell-wall ultrastructure and lacks classical cell-wall lipopolysaccharide // *J Med Microbiol.* 1989. Vol. 29, N 3. P. 229–235. doi: 10.1099/00222615-29-3-229
23. Scott T.G., Curran B., Smyth C.J. Electron microscopy of adhesive interactions between *Gardnerella vaginalis* and vaginal epithelial cells, McCoy cells and human red blood cells // *J Gen Microbiol.* 1989. Vol. 135, N 3. P. 475–480. doi: 10.1099/00221287-135-3-475
24. Taylor-Robinson D. The bacteriology of *Gardnerella vaginalis* // *Scand J Urol Nephrol Suppl.* 1984. Vol. 86. P. 41–55.
25. Ильина Т.С., Романова Ю.М. Бактериальные биоплёнки: роль в хронических инфекционных процессах и поиск средств борьбы с ними // *Молекулярная генетика, микробиология и вирусология*. 2021. Т. 39, № 2. С. 14–24. EDN: RHLJAM doi: 10.17116/molgen20213902114
26. Хрянин А.А. Биоплёнки микроорганизмов: современные представления // *Антибиотики и химиотерапия*. 2020. Vol. 65, N 5–6. P. 70–77. EDN: NQITOE doi: 10.37489/0235-2990-2020-65-5-6-70-77
27. Jung H.S., Ehlers M.M., Lombaard H., et al. Etiology of bacterial vaginosis and polymicrobial biofilm formation // *Crit Rev Microbiol.* 2017. Vol. 43, N 6. P. 651–667. doi: 10.1080/1040841X.2017.1291579
28. Пестрикова Т.Ю., Юрасова Е.А., Котельникова А.В., и др. Современный подход к лечению рецидивирующего бактериального вагиноза у женщин репродуктивного периода // *Гинекология*. 2018. Т. 20, № 2. С. 55–58. EDN: XTGRVB doi: 10.26442/2079-5696_2018.2.55-58
29. Nickel J.C., Ruseska I., Wright J.B., Costerton J.W. Tobramycin resistance of cells of *Pseudomonas aeruginosa* growing as a biofilm on urinary catheter material // *Antimicrob Agents Chemother.* 1985. Vol. 27, N 4. P. 619–624. doi: 10.1128/AAC.27.4.619
30. Березовская Е.С., Макаров И.О., Гомберг М.А., и др. Биоплёнки при бактериальном вагинозе // *Акушерство, гинекология и репродукция*. 2013. Т. 7, № 2. С. 34–36. EDN: RRPOER
31. Simões M., Simões L.C., Vieira M.J. Species association increases biofilm resistance to chemical and mechanical treatments // *Water Res.* 2009. Vol. 43, N 1. P. 229–237. doi: 10.1016/j.watres.2008.10.010
32. Khan J., Tarar S.M., Gul I., et al. Challenges of antibiotic resistance biofilms and potential combating strategies: a review // *3 Biotech.* 2021. Vol. 11, N 4. P. 169. doi: 10.1007/s13205-021-02707-w
33. Michaelis C., Grohmann E. Horizontal gene transfer of antibiotic resistance genes in biofilms // *Antibiotics (Basel)*. 2023. Vol. 12, N 2. P. 328. doi: 10.3390/antibiotics12020328
34. Bonnardel F., Haslam S.M., Dell A., et al. Proteome-wide prediction of bacterial carbohydrate-binding proteins as a tool for understanding commensal and pathogen colonisation of the vaginal microbiome // *NPJ Biofilms Microbiomes*. 2021. Vol. 7, N 1. P. 49. doi: 10.1038/s41522-021-00220-9
35. Marín E., Haesaert A., Padilla L., et al. Unraveling *Gardnerella vaginalis* surface proteins using cell shaving proteomics // *Front Microbiol.* 2018. Vol. 9. P. 975. doi: 10.3389/fmicb.2018.00975
36. Hardy L., Jespers V., Abdellati S., De Baetselier I. A fruitful alliance: the synergy between *Atopobium vaginae* and *Gardnerella vaginalis* in bacterial vaginosis-associated biofilm // *Sex Transm Infect.* 2016. Vol. 92, N 7. P. 487–491. doi: 10.1136/sextrans-2015-052475
37. Castro J., Machado D., Cerca N. Unveiling the role of *Gardnerella vaginalis* in polymicrobial Bacterial Vaginosis biofilms: the impact of other vaginal pathogens living as neighbors // *ISME J.* 2019. Vol. 13, N 5. P. 1306–1317. doi: 10.1038/s41396-018-0337-0
38. Castro J., Cerca N. BV and non-BV associated *Gardnerella vaginalis* establish similar synergistic interactions with other BV-associated microorganisms in dual-species biofilms // *Anaerobe*. 2015. Vol. 36. P. 56–59. doi: 10.1016/j.anaerobe.2015.10.008
39. Schwebke J.R., Muzny C.A., Josey W.E. Role of *Gardnerella vaginalis* in the pathogenesis of bacterial vaginosis: a conceptual model // *J Infect Dis.* 2014. Vol. 210, N 3. P. 338–343. doi: 10.1093/infdis/jiu089
40. Shvartsman E., Hill J.E., Sandstrom P., MacDonald K.S. *Gardnerella* revisited: species heterogeneity, virulence factors, mucosal immune responses, and contributions to bacterial vaginosis // *Infect Immun.* 2023. Vol. 91, N 5. P. e0039022. doi: 10.1128/iai.00390-22
41. Coudray M.S., Madhivanan P. Bacterial vaginosis — A brief synopsis of the literature // *Eur J Obstet Gynecol Reprod Biol.* 2020. Vol. 245. P. 143–148. doi: 10.1016/j.ejogrb.2019.12.035
42. Абатуров А.Е. Полисахаридразрушающие ферменты как агенты, диспергирующие бактериальные биоплёнки // *Здоровье ребёнка*. 2020. Т. 15, № 4. С. 271–278. EDN: WKPGMH doi: 10.22141/2224-0551.15.4.2020.208478
43. Marshall A.O. Managing recurrent bacterial vaginosis: insights for busy providers // *Sex Med Rev.* 2015. Vol. 3, N 2. P. 88–92. doi: 10.1002/smrj.45

AUTHORS' INFO

***Kseniya A. Rossolovskaya**, Graduate Student;
address: 8 build. 2, Trubetskaya Str., 119991 Moscow, Russia;
ORCID: 0000-0002-7026-1607;
eLibrary SPIN: 4432-5748;
e-mail: dr.rossolovskaya@yandex.ru

Natalia S. Trifonova, MD, Dr. Sci. (Medicine), Professor;
ORCID: 0000-0002-2891-3421;
eLibrary SPIN: 4753-5430;
e-mail: trifonova.nataly@mail.ru

Irina V. Gadaeva, MD, Cand. Sci. (Medicine);
ORCID: 0000-0003-0144-4984;
eLibrary SPIN: 9593-1990;
e-mail: gadaeva-gin@gmail.com

Leonid G. Spivak, MD, Dr. Sci. (Medicine), Professor;
ORCID: 0000-0003-1575-6268;
eLibrary SPIN: 5230-8811;
e-mail: leonid.spivak@gmail.com

ОБ АВТОРАХ

***Россоловская Ксения Антоновна**, аспирант;
адрес: Россия, 119991, Москва, ул. Трубецкая, д. 8, стр. 2;
ORCID: 0000-0002-7026-1607;
eLibrary SPIN: 4432-5748;
e-mail: dr.rossolovskaya@yandex.ru

Трифорова Наталья Сяитовна, д-р мед. наук, профессор;
ORCID: 0000-0002-2891-3421;
eLibrary SPIN: 4753-5430;
e-mail: trifonova.nataly@mail.ru

Гадаева Ирина Викторовна, канд. мед. наук;
ORCID: 0000-0003-0144-4984;
eLibrary SPIN: 9593-1990;
e-mail: gadaeva-gin@gmail.com

Спивак Леонид Григорьевич, д-р мед. наук, профессор;
ORCID: 0000-0003-1575-6268;
eLibrary SPIN: 5230-8811;
e-mail: leonid.spivak@gmail.com

* Corresponding author / Автор, ответственный за переписку